



Publication number:

0 050 424
A1

12

EUROPEAN PATENT APPLICATION

21 Application number: 81304411.2

51 Int. Cl.: G 01 N 33/54, G 01 N 27/26

22 Date of filing: 24.09.81

30 Priority: 24.09.80 US 190361
02.09.81 US 297962

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43 Date of publication of application: 28.04.82
Bulletin 82/17

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84 Designated Contracting States: AT BE CH DE FR GB IT
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54 Method for producing an analytical antibody probe, an analytical antibody probe, and a method for analysing a sample for certain antibodies.

57 The complexity of antigenic test material in tests which employ the antigen-antibody reaction increases the likelihood of false results. In particular, the complexity of antigenic test material in diagnostic techniques increases the likelihood of false positive signals generated due to antibodies stimulated by organisms showing some but all antigenic properties with the test material. In conventional tests, a single positive or negative signal is obtained which is the result of a complex and undifferentiated array of structural components of an organism or material antigenically related to it. In contrast, the present invention greatly expands the use of the antigen-antibody reaction in analysis techniques and diagnosis by providing a multi-signal.

analysing a sample to determine the presence and distribution of antibodies specific for particular protein and/or polysaccharide material.

The invention provides a method for producing an analytical antibody probe for detecting the presence of antibodies specific for particular protein and/or polysaccharide material, which method comprises disposing upon a solid-state substrate in a pre-determined spatial relationship a set of differentiated antigenic components which have been derived from said protein or polysaccharide material, said set of differentiated antigenic components being selected such that at least a subset thereof is known to be reactive with said antibodies. The invention also embraces the analytical antibody probe as such and a corresponding method for

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METHOD FOR PRODUCING AN ANALYTICAL ANTIBODY PROBE,
AN ANALYTICAL ANTIBODY PROBE, AND A METHOD FOR
ANALYSING A SAMPLE FOR CERTAIN ANTIBODIES

The present invention provides, in general, a method
5 for producing an analytical antibody probe, an analytical
antibody probe, and a method for analysing a sample for
certain antibodies. The invention has particular value in
the diagnosis of disease or allergy in patients. More
particularly, the invention permits an improved method for
10 such diagnosis, and provides an improved product for
conducting such diagnosis.

Many clinical diagnostic techniques have, as their
fundamental basis, the antigen-antibody reaction. This
reaction serves as a defence against micro-organisms and
15 other foreign bodies as part of the body's normal immuno-
logical response. Detection of the presence of antigen-
antibody reactions in tests performed upon serum obtained
from a patient may indicate the presence or absence of
antibodies in the patient's serum. A positive test for
20 reaction of antibody with a specific antigen may indicate
a presence of a corresponding disease or at least suggest
that diagnostic conclusion.

Known clinical diagnostic procedures that test for the
antigen-antibody reaction in sera (serotests) may take a
25 wide variety of forms. Some utilize antigens anchored to
the surface of inert particles. In the presence of specific
antibody, these particulate antigens clump visibly in an
agglutination reaction. Such a procedure is widely employed
in the diagnosis of syphilis and is known as the Venereal
30 Disease Research Laboratory (VDRL) test. Other tests may
involve the attachment of a fluorescent or radioactive
moiety in such a way that its presence indicates that the
antigen-antibody reaction has occurred. In a similar manner,
enzymes have been linked as a detectable moiety - the so-
35 called enzyme linked immunosorbent assay (ELISA).

The hallmark of conventional serotests is that they measure a single positive or negative signal. Signals may be, for example, hemagglutination, hemagglutination inhibition, complement fixation, surface fluorescence, particle
5 agglutination and so on. That single positive or negative signal is usually a complex and undifferentiated array of the structural components of an organism or material antigenically related to it. This complexity of the antigen test material increases the likelihood that false positive
10 signals will be generated due to antibodies stimulated by other organisms sharing some but not all antigenic properties with the test material. In contrast the present invention enables the provision of a diagnostic method and means by which the antigen-antibody reaction may be greatly expanded
15 as a diagnostic tool. Using the invention the presence of a specific disease, a specific stage of a disease, or an allergy in a patient may be detected by a simple procedure.

Thus the invention provides a method for producing an analytical antibody probe for detecting the presence of
20 antibodies specific for particular protein and/or polysaccharide material, which method comprises disposing upon a solid-state substrate in a pre-determined spatial relationship a set of differentiated antigenic components which have been derived from said protein or polysaccharide
25 material, said set of differentiated antigenic components being selected such that at least a subset thereof is known to be reactive with said antibodies. The invention also provides an analytical antibody probe for detecting the presence of antibodies specific for particular protein and/
30 or polysaccharide material, which probe comprises a solid-state substrate upon which is disposed in a pre-determined spatial relationship a set of differentiated antigenic components derived from said protein or polysaccharide material, said set of differentiated antigenic components
35 being selected such that at least a subset thereof is known

to be reactive with said antibodies.

Specific aspects of the invention are:-

(a) a method for producing a diagnostic antibody probe, comprising, disposing upon a solid-state substrate a set of differentiated antigenic components of a larger set of proteins or polysaccharides in a predetermined spatial relationship, said larger set being related to a disease, disease stage or allergy under diagnosis, said set of differentiated antigenic components being selected such that at least a subset of said differentiated antigenic components is known to be reactive with antibodies present in serum obtained from a patient having the specific disease stage or allergy; and

(b) a diagnostic antibody probe comprising a solid-state substrate upon which is disposed in a pre-determined spatial relationship a set of differentiated antigenic components of a larger set of proteins or polysaccharides, said larger set being related to a disease, disease stage or allergy under diagnosis, said set of differentiated antigenic components being selected such that at least a subset of said differentiated antigenic components is known to be reactive with antibodies present in serum obtained from a patient having the specific disease, disease stage or allergy.

One further aspect of the invention is a method for analysing a sample to determine the presence and distribution of antibodies specific for particular protein and/or polysaccharide material, which method comprises providing a solid-state substrate upon which is disposed in a pre-determined spatial relationship a set of differentiated antigenic components derived from said protein or polysaccharide material, said set of differentiated antigenic components being selected such that at least a subset thereof is known to be reactive with said antibodies, contacting said substrate with said sample under conditions which permit reaction of any of said antibodies in the sample with the

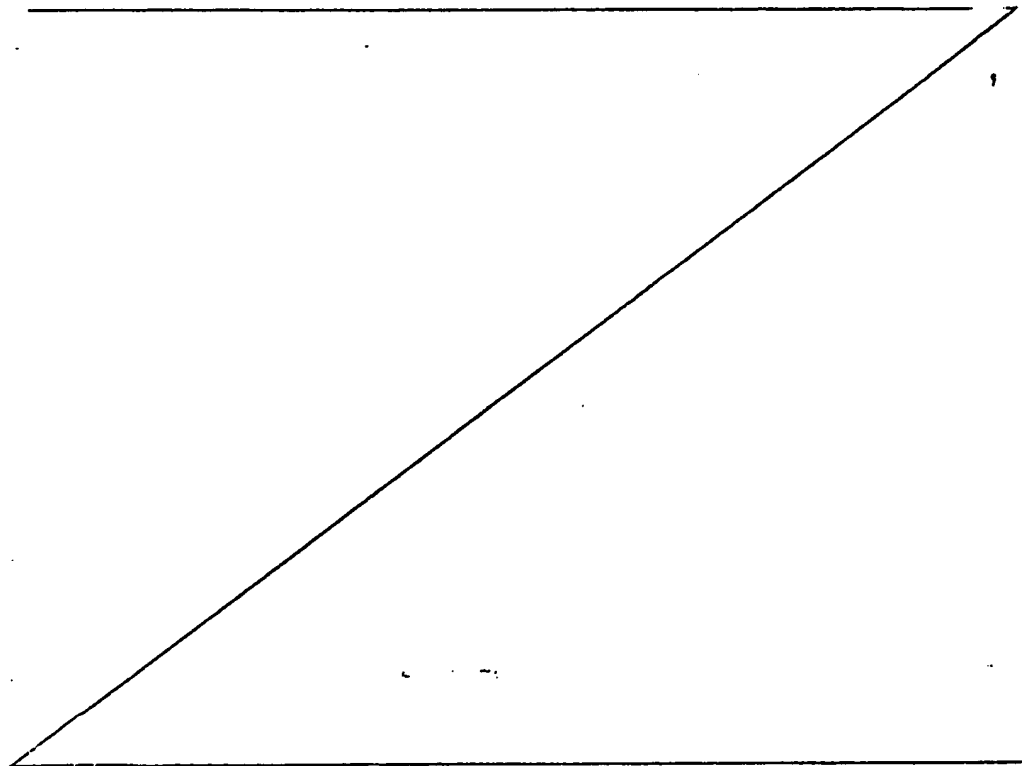
antigenic components on the substrate, and detecting the existence and pattern of antigen-antibody reactions on the substrate.

The invention will now be further described with reference to the accompanying drawings, in which:-

FIGURE 1 is a photograph of a conventional SDS polyacrylamide gel showing the total protein profile of T. pallidum after staining;

10 FIGURE 2 is a photograph of an autoradiogram of twelve diagnostic strips of T. pallidum, constructed in accordance with the invention, after exposure to sera drawn from patients, eight of whom had different stages of syphilis and four of whom had exhibited false positive
15 (BFP) on standard serological tests for syphilis;

FIGURE 3 is a photograph of an autoradiogram of nine diagnostic strips of T. pallidum, constructed in



accordance with the invention, after exposure to sera drawn from patients suffering from late syphilis and secondary syphilis;

5 FIGURE 4 is a photograph of an autoradiogram of ten diagnostic strips of T. pallidum, constructed in accordance with the invention, after exposure to sera drawn from patients suffering from early latent and late latent syphilis;

10 FIGURE 5 is a photograph of conventional SDS gels showing the total protein profiles of five different immunotypes of Chlamydia;

15 FIGURE 6 is a photograph of an autoradiogram of five diagnostic strips of Chlamydia trachomatis, constructed in accordance with the invention, after exposure to rabbit antiserum to Chlamydia trachomatis immuno type B;

 FIGURE 7 is a photograph of an autoradiogram of seven diagnostic strips of Toxoplasma gondii, constructed in accordance with the invention, after exposure to sera drawn from patients with chronic and acute toxoplasmosis;

20 FIGURE 8 is a photograph of an autoradiogram of diagnostic strips of cytomegalovirus infected cells and cell free preparation of partially purified virus, constructed in accordance with the invention, after exposure to patient sera;

25 FIGURE 9 is a photograph of an autoradiogram of diagnostic strips of rabbit kidney cells infected with Herpes simplex virus I or II, constructed in accordance with the invention, after exposure to sera drawn from patients suffering from Herpes infections;

30 FIGURE 10 is a photograph of an autoradiogram of diagnostic strips of capsular polysaccharide types 4 and 8 for Streptococcus pneumoniae, constructed in accordance with the invention, after exposure to an immune serum;

35 FIGURE 11 is a photograph of an autoradiogram of a diagnostic strip of bee venom, constructed in accordance

with the invention, after exposure to serum drawn from a patient allergic to bee venom;

5 FIGURES 12 and 13 are photographs of an autoradiogram of diagnostic strips of type L₂ C. trachomatis, constructed in accordance with the invention, after reaction with sera from patients with type F infection and type D infection, respectively;

10 FIGURE 14 is a photograph of an autoradiogram of diagnostic strips of T. pallidum, constructed in accordance with the invention, after exposure to sera drawn from patients suffering from syphilis and after exposure to IgG and IgM specific antisera probes;

15 FIGURE 15 is a photograph of an autoradiogram of diagnostic strips of Toxoplasma gondii, constructed in accordance with the invention, after exposure to serum drawn from a patient suffering from acute toxoplasmosis and after exposure to IgG and IgM specific probes;

20 FIGURE 16 is a photograph of an autoradiogram of diagnostic strips of cytomegalovirus, constructed in accordance with the invention, after exposure to a serum pool drawn from patients suffering from viral infection and after exposure to IgG and IgM specific probes;

25 FIGURE 17 is a photograph of an autoradiogram of diagnostic strips of Toxoplasma gondii, constructed in accordance with the invention, when the Toxoplasma antigens were separated on a nondenaturing isoelectric focusing gel;

30 FIGURE 18 is a photograph of an autoradiogram of diagnostic strips of Toxoplasma gondii, prepared in accordance with the invention, when the Toxoplasma antigens are separated on a nondenaturing native gel system; and

 FIGURE 19 is a photograph of an autoradiogram of a diagnostic strip of cloned T. pallidum - specific antigens, constructed in accordance with the invention, after exposure to sera from patients suffering from syphilis.

35 Very generally, the use of the invention for diagnosis involves a method com-

prising providing a solid-state substrate upon which is disposed in a predetermined relationship a set of differentiated antigenic components of a larger set of proteins or polysaccharides. The larger set is related to the disease or allergy under diagnosis. The set of components is selected such that at least a subset of the components is known to be reactive with antibodies present in serum obtained from a patient having the specific disease or allergy. The substrate is contacted with serum obtained from the patient under diagnosis under conditions which permit reaction of antibodies in the serum with antigenic components on the substrate. The existence and pattern of antigen-antibody reactions on the substrate is then detected for correspondence with the pattern of those components on the substrate known to be reactive for the specific disease, the specific stage of the disease, or the allergy.

The method detailed here for disease diagnosis differs markedly from conventional serotests in that what comprises a "positive" test is not the presence of one signal, but rather the appearance of a series of signals that represent antibody response to defined antigens associated with a specific pathogen or disease. The number and nature of the signals which define a disease state are defined empirically for each specific disease; all humans with this disease display the same multiple signals characteristic of that disease.

More particularly, the invention, although based upon the antigen-antibody reaction phenomenon, take advantage of a discovery of major significance. If protein or polysaccharide material related to a specific disease, or stage of a disease, or an allergy is differentiated in such a way as to establish a spatial relationship of differentiated antigenic components, a plurality of antigen-antibody reactions may be detected upon exposure to serum

from a diseased patient. The extent and pattern of such reactions, with proper differentiation, can be made highly specific to a given disease. Accordingly, a sort of molecular antigenic "fingerprint" may be established which will identify a particular disease and no other. Moreover, this molecular antigenic "fingerprint" may be designed so as to distinguish between historical antibodies, i.e., residual IgG antibodies which indicate previous exposure to a given antigen, and current antibody, i.e., recent IgM antibodies which indicate very recent exposure to a given antigen. In the case of allergy diagnosis, the antibody detected is immunoglobulin IgE.

The method of the invention utilizes a solid-state substrate upon which is disposed in a predetermined spatial relationship a set of differentiated antigenic components of a larger set of proteins or polysaccharides. This larger group is related to the disease, the stage of the disease, or the allergy under diagnosis. It may be the pathogen itself (e.g., Herpes virus I or II0 or it may be a tissue sample (e.g., of a sarcoma tumor) or it may be an allergen such as various types of grasses or pollen. Frequently, the source of the larger set of proteins or polysaccharides will itself be antigenic for a particular disease but, because of non-specificity, cross reactivity, or other problems, is unsatisfactory for diagnostic use.

The set of differentiated antigenic components of the larger set of proteins or polysaccharides is chosen for its specific ability to identify the disease, the stage of the disease, or allergy of interest. These components are selected such that at least a subset of the components is known to be reactive with antibodies present in serum obtained from a patient having the specific disease or allergy. The subsets are determined empirically in accordance with techniques described below. The process by which the components are differentiated depends upon the

particular disease, the larger protein or polysaccharide from which the components are derived, and the particular antibodies which are to be detected (e.g., IgG, IgM, or IgE). Such differentiation processes may include, but are not limited to, electrophoresis (SDS or native gel), isoelectric focusing, thin layer chromatography, and centrifugation. Another differentiation process, once the nature of the subject of components is ascertained, is to produce each component of the subset separately by means of genetically modified microorganisms. Each component may then be placed separately on the substrate.

In any case, the individual differentiated components are positioned upon a solid-state substrate such as a cellulose strip. The precise manner of attaching the components to the substrate will depend upon the nature of the components and the substrate. For example, if electrophoresis is utilized as a differentiation process, a useful transfer technique is the so-called filter affinity transfer as described by Erlich, H.A., et al., in Journ. Biol. Chem., 254:12240-12247 (1979).

A typical solid-state substrate may be a cellulose strip to which a plurality of differentiated components of the pathogen responsible for a disease have been transferred. When the strip is exposed to a patient's serum, components which are reactive with antibodies in the serum bind to it and may be detected by any suitable assay. Before such exposure, the strip may be blank in appearance with the differentiated components not visible. Once exposed to antibodies, the antibody-antigen reactions cause the antibodies to bind to the strip in a pattern that is indicative of the present or absence of a specific disease. Through previous empirical testing, it can be readily established as to which differentiated proteins or groups of proteins on a given substrate will react with antibodies in the serum of a patient having the specific

disease that is of interest. Thus, separate strips may be produced that are specific to, for example, Chlamydia, syphilis, and gonorrhoea, respectively.

5 Once the strip or solid-state substrate is produced with the empirically determined and selected pattern of differentiated antigenic components established on the substrate, it is in a form useful for clinical diagnosis. For such a use, the substrate is exposed or contacted with serum obtained from the patient under diagnosis. The conditions under which the contact occurs are established so as to permit reaction of antibodies in the serum with antigenic components on the substrate.

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 Following suitable illumination steps, e.g., radioactively labelled probes specific for human immunoglobulin classes and autoradiography, the strip is examined to ascertain the pattern, if any, of antibody-antigen reactions which has developed. If the pattern corresponds to the pattern known for the particular disease to which the strip or substrate corresponds, a positive diagnosis is obtained. Otherwise, the diagnosis is negative. Actual detection of the antigen-antibody reactions may employ other than autoradiographic assay of the type shown in FIGURE 2. For example, colorimetric assays may also be employed.

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25 Traditionally, radioactively labelled S. aureus protein A has been used as a probe for IgG antibodies. To distinguish between reactions with historical antibody (IgG) and new antibody (IgM), antiserum to human IgM may be labelled in a variety of ways, e.g., ¹²⁵I or fluorescein, and used as probe to detect formation of new human antibody (IgM). This provides the ability to distinguish between persons with a history of a disease who do not currently have an active form of that disease and persons with the active form of the disease. Historical antibodies will, in many cases, remain with a cured person

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for life.

The present invention will be more readily understood by means of the following examples. These examples are set forth for the purpose of elaborating on the invention and are not intended to limit the invention in any way.

Example I. Use of the Invention for the Correct Diagnosis of Syphilis

FIGURE 1 is a photograph showing the total protein profile of Treponema pallidum. These proteins are separated and stained on a conventional SDS polyacrylamide gel. To obtain this profile, intact T. pallidum was suspended in an electrophoresis sample buffer comprised of 62.5 mM tris (pH 6.8), 2% sodium dodecylsulfate, and 5% mercaptoethanol. The sample was then applied to an SDS polyacrylamide gel system as described in Laemmli, U.K., Nature (London) 227:680-685 (1970). The gel was run until the tracking dye reached the bottom of the gel.

In FIGURE 1, the left-hand column represents the T. pallidum profile, whereas the right-hand column is a system of molecular weight markers as is well known in the use of polyacrylamide gel separations. FIGURE 1 provides a base for comparison of the actual protein separation with the antigenic activity described in FIGURE 2.

In FIGURE 2, diagnostic strips were prepared in accordance with the invention using the total protein separation of T. pallidum illustrated in FIGURE 1. To prepare the strips, the gel is overlaid with nitrocellulose paper as described by Towbin, H., Staehlin, T., and Gordon, J., PNAS (USA) 76:4350-4354 (1979). The paper is then covered with scouring pads and supported by lucite grids with numerous pores. The assembly is held together with rubber bands and is then placed in a single chamber for electrophoresis such that the surface of the gel applied directly to the paper is facing the anode. Elec-

trophoresis is performed in an elctrode buffer comprised of 25 mM tris, 192 mM glycine, and 20% volume/volume methanol at pH 8.3. Electrophoresis is carried out for 90 minutes. The nitrocellulose at the end of this procedure contains the proteins arrayed as they have been separated according to molecular weight and is referred to as the blot.

The blots were then soaked in a solution of 1% bovine serum albumin in a buffer comprised of 50 mM tris (pH 7.5), 0.9% sodium chloride, 0.25% gelatin, 0.2% sodium azide, and 0.1% NP 40 (TSGAN) for ten minutes at room temperature. This was to saturate all remaining reactive sites on the paper. At this point, the blots are ready for use and may be stored by freezing or other suitable means.

Each of the strips in FIGURE 2 is a blot of the T. pallidum total protein profile after exposure to patient serum representing different stages of syphilis (8 patients) and representing non-syphilitic patients who showed false positive (BFP) in standard serological tests for syphilis (4 patients). Serum dilutions were used at 1:1000 with twelve hours at room temperature with gentle shaking. After this period of incubation with serum, the blots were rinsed several times with TSGAN and then washed with TSGAN for 20-60 minutes at room temperature, again with gentle agitation. Then 2-4 microcuries of protein A with a specific activity of greater than 10^7 counts per minute/microgram is added in a volume of 100-200 milliliters of TSGAN and incubation continued with a gentle agitation for 60 minutes at room temperature. A similar incubation buffer system is described in Renard, J., Reiser, J., and Stark, G. R., PNAS (USA) 76: 3116-312 (1979).

The blots were then rinsed several times with TSGAN, washed with TSGAN with gentle agitation at room temperature for 20 minutes and then rinsed several times

FIGURE 5, left-hand columns 1-5, show the stained total proteins of five different immunotypes of Chlamydia. Column 6 in FIGURE 5 is a molecular weight marker system. By standard serology, these immunotypes are non-crossreactive. Accordingly, a separate antiserum for clinical use must be prepared for each serotype.

FIGURE 6 shows the strips of the invention prepared in accordance with Example I after exposure to rabbit antiserum to Chlamydia trachomatis immuno type B and autoradiography. Preparations and procedures were as in Example I. All C. trachomatis immuno types have extensive cross-reaction of the major antigenic proteins. It may be seen that the left-hand 4 strips show strong reaction whereas the strip in the far right-hand side, specific for C. psittaci, shows weak relatedness of only two antigens. This illustrates that a single C. trachomatis immunotype is a sufficient source of antigens for testing human infection with any other C. trachomatis immunotypes, and yet provides specificity in that other types of Chlamydia may be readily distinguished.

Example III. Use of the Invention To Detect Toxoplasma gondii Antigen

FIGURE 7 is a photograph showing various antigenic bands of Toxoplasma gondii that react with antibodies in the sera of patients with chronic and acute toxoplasmosis. To obtain the observed patterns, the toxoplasmal antigens were separated on a conventional SDS polyacrylamide gel. A sonicate of Toxoplasma was suspended in an equal volume of electrophoresis sample buffer consisting of 0.125M trizma base (pH 6.8), 2.5% sodium dodecylsulfate, and 2.5% β -mercaptoethanol. The sample was then applied to an SDS polyacrylamide gel system as described in Laemmli, U. K., Nature (London) 227:680-685 (1970). The gel was run until the tracking dye reached the bottom of the gel.

The gel was then washed for 15 minutes in water and in two 5-minute washes of 50 mM sodium acetate, pH 7.0. The peptide components of Toxoplasma, separated by molecular weight in the SDS polyacrylamide gel, were electrophoretically transferred onto cyanogen bromide activated paper as follows. The gel was placed on a scotch bright pad covered with filter paper. A sheet of cyanogen bromide treated filter paper was laid on the gel and another sheet of filter paper and a scotch bright pad was placed on top. The assembly is placed in an E-C electroblot unit with the cyanogen bromide paper facing the anode. Electrophoresis was carried out in 50 mM sodium acetate, pH 7, at 25 volts for one hour.

All remaining reactive sites on the cyanogen bromide paper are bound and/or inactivated by soaking the paper in a solution of 1M glycine and 1% bovine serum albumin for 0.5 to 3 hours. The paper was washed three times for 5-15 minutes each with agitation in a wash solution containing 0.1% ovalbumin, 0.1% tween 20, 0.02% sodium azide in phosphate buffered saline. The paper is then incubated at room temperature with gentle agitation for 2-3 hours in diluted human serum. The sera used in FIGURE 7 are from patients with chronic or acute toxoplasmosis. The patient's serum is diluted 1:25 in wash solution.

After the incubation with serum, the paper is washed three times for 5-15 minutes with shaking in wash solution. The ¹²⁵I Protein A is added to the paper using a 1:200 dilution of stock (~5µg/ml, 15µci/µg) in wash solution. The protein A is iodinated using the chloramine T method as described by Erlich, H., Cohen, S., and McDevitt, H., in Cell, 13:681-689 (1978). The paper is incubated with the ¹²⁵I Protein A for 1-3 hours at room temperature with agitation. The paper is again washed as above, dried and placed under Kodak XAR-5 X-ray film for 16 hours.

infected cells were separated on 9.5% denaturing SDS polyacrylamide gels as described in Example I. Diagnostic strips derived from the gels were then cross-reacted with serum from patients suffering with type I or type II Herpes infections. FIGURE 9 is a photograph of an autoradiogram of these diagnostic strips following exposure to appropriate radioactive probes.

Strip 1, i.e., the far right-hand strip on the photograph, shows a standard system of molecular weight markers. Strips 2 and 3 show how serum from patient J.K. reacts with antigens from the two types of Herpes. Strip 2 represents proteins derived from Herpes virus type I while strip 3 represents proteins derived from Herpes virus type II. Comparison of the strips shows that serum from patient J.K. contains IgG antibodies that react strongly with the peptides derived from Herpes virus type I infected cells, and only weakly with peptides derived from Herpes virus type II infected cells.

Strips 8, 9 and 10 show the reaction of serum from patient L.O. Strips 8 and 9 represent proteins derived from Herpes virus type I; strip 8 proteins demonstrate an IgG reaction while strip 9 demonstrates the presence of IgM's. Strip 10 represents proteins derived from Herpes virus type II. Comparison of the strips shows that serum from patient L.O. contains IgG antibodies to proteins derived from Herpes virus type II infected cells. They also show L.O.'s serum reacts only weakly with protein derived from Herpes virus type I infected cells. Strips 4-7 and strips 11 and 12 are not relevant to this Example.

Example VI. Use of the Invention for
Detection of Polysaccharide Antigen

FIGURE 10 illustrates the reaction of immune serum with pneumococcal capsular polysaccharide of types 4 and 8. Three micrograms of purified polysaccharide were

and stained on SDS polyacrylamide gel as is described in Example I. Rabbit antiserum to human IgM was labelled with ^{125}I and used as a probe for IgM. ^{125}I labelled Protein A of S. aureus was used as a probe for IgG.

5 The furthest right-hand column in the FIGURE 14 -
photograph is a standard system of molecular weight markers
well known to those skilled in the art. The remaining six-
teen columns represent diagnostic strips after exposure to
serum from eight different patients. The right-hand strip
10 (A) in each pair was probed with ^{125}I -rabbit anti-human IgM
and the left-hand strip (B) with ^{125}I -Protein A, which is
specific for IgG. The first five patients (i.e., the first
ten strips) illustrate sera from patients with primary syph-
15 ilis. The next three patients (i.e., the remaining six
strips) illustrate sera from normal humans not infected with
syphilis. Strips from these last three patients show that
uninfected humans have little or no IgG or IgM antibodies
to antigenic proteins derived from the organism that causes
20 syphilis. In contrast, all patients with primary syphilis
have IgG and IgM antibodies to the proteins from T. pallidum.
The IgG or historic antibodies are clearly distinguishable
from the current IgM antibodies in all the patients with
primary syphilis.

FIGURE 15 shows that the diagnostic strips of the
25 present invention can be used to distinguish between IgG and
IgM antibodies in the sera of patients suffering from toxo-
plasmosis. Toxoplasma gondii were separated and run on SDS
polyacrylamide gel, as is described in Example III; transfer
of the gel protein pattern to the diagnostic strips is also
30 described in that Example. Strips 1 and 2 were incubated
with the serum from a patient with an acute case of toxoplas-
mosis. ^{125}I labelled Protein A of S. aureus was used as a
probe for IgG in strip 1. ^{125}I labelled affinity-purified
Goat antibodies to human IgM was used as a probe for IgM.
35 Strip 3 represents well known molecular markers.

FIGURE 16 illustrates that the diagnostic strips can be used to distinguish between IgG and IgM antibodies in the sera of patients infected with cytomegalovirus. The samples and steps were prepared as outlined in Examples III and IV. Again, ^{125}I labelled Protein A was used as a probe for IgG and ^{125}I labelled affinity-purified Goat antibodies to human IgM were used as a probe for IgM. Strip 1 in FIGURE 16 shows the presence of IgG, strip 2 the presence of IgM. Strip 3 is a system of standard molecular markers.

10 Example X. Antigenic Proteins for the Diagnostic Strips Can be Separated on Nondenaturing Gels.

A. Use of nondenaturing isoelectricfocusing gel.

15 FIGURE 17 illustrates the Toxoplasma antigen bands observed when the antigen is separated on nondenaturing isoelectricfocusing gel and sequentially incubated with patient sera and ^{125}I Protein A. A sonicate of Toxoplasma gondii is made 1% in noniodet P40. Nonsolubilized membranes are pelleted by centrifugation at 15,000 RPM for 2 minutes. The supernate is pipetted directly onto the pre-run gel.

20 The gel is made 5% in acrylamide, 0.0013% in bis acrylamide (T=5.1%, C=2.6%), 13% in sucrose, 2% noniodet P40, and 5% in ampholytes pH 3.5-10.0. The gel is polymerized with ammonium persulfate and TEMED for 1 hour. The gel is prerun for 1-2 hours of 30ma constant current with a voltage maximum of 1000. The anode solution is 1M phosphoric acid, the cathode 1M sodium hydroxide. The samples are added to the gel and electrophoresed for 2.5 hours at 1000 volts. The separated antigens are transferred to cyanogen bromide treated paper as outlined in Example III, except that the gel is not washed with water and sodium acetate before transfer.

30 In FIGURE 17, strips 1-4 show the isoelectric bands from patients suffering from toxoplasmosis. Strips 5 and 6 are from uninfected humans and therefore show no bands. Strip 7 is a positive with a rabbit antiserum.

B. Use of nondenaturing native gel.

FIGURE 18 shows the Toxoplasma gondii banding pattern obtained when antigens are separated on a non-denaturing native gel system and sequentially incubated with patient sera and ¹²⁵I Protein A. The procedure described in Example III for electrophoresis and transfer of Toxoplasma is utilized with the following modifications. The gel is made 7.5% in acrylamide, 0.2% in bis acrylamide, 2% in noniodet P40, and 75mM trizma base plus 32mM boric acid pH 8.9. The gel is polymerized with ammonium persulfate and TEMED for 20 minutes. The gel is overlaid with a stacking gel made 4% in acrylamide, 0.1% in bis acrylamide, 2% noniodet P40, and 37.5mM trizma base plus 16mM boric acid pH 8.9. This stacking gel is polymerized with ammonium persulfate and TEMED for 10 minutes. The electrode buffer for the system is 0.1% noniodet P40, 75mM trizma base, 32mM boric acid pH 8.9. The Toxoplasma sonicated organisms are made 1% in noniodet P40 and applied to the gel as in Example III.

Strip 1 is from an uninfected patient and therefore shows no bands characteristic of Toxoplasma antigens. Strips 2-6 are from patients suffering from various forms of toxoplasmosis; all show bands characteristic of the disease. Strip 7 is a positive control with a rabbit anti-serum.

Example XI. The Antigenic Proteins of the Invention can be Produced by Genetically Engineered Microorganisms.

The antigenic proteins used in the invention can be products of genes derived from antigenic organisms that have been separately cloned into suitable genetically engineered host microorganisms. Expression of cloned T. pallidum DNA in E. coli illustrates such antigenic protein production.

In this Example, Treponema pallidum were first harvested from the testicles of ten rabbits. The testicles

were extensively minced in phosphate-buffered saline before the resulting extract was subjected to several cycles of differential centrifugation to remove cellular debris. The final supernatant, which contained motile and virulent T. pallidum, was further purified on a density gradient using a homogenous solution of Percoll, produced by Pharmacia Corporation, Piscataway, New Jersey 08854. Centrifugation at 20,000 RPM for 20 minutes produced a band of relatively pure, motile and virulent T. pallidum. The band was pulled from the Percoll gradient material, subjected to a dilution in phosphate-buffered saline, and then pelleted by ultracentrifugation at 100,000 x G for 2 hours. The pellet of T. pallidum was resuspended in buffer containing tris-EDTA, pH 7.5, before treatment with the detergent Sarcosyl, (N-lauroylsarcosine) produced by Sigma Chemicals, St. Louis, Missouri 63178, to liberate the treponemal DNA. The resulting DNA-detergent extract was centrifuged to equilibrium on a cesium chloride density gradient. The treponemal DNA band was then pulled from the gradient and dialysed against Sau3A I restriction buffer minus magnesium. The dialyzed DNA was partially digested with Sau3A I restriction endonuclease using techniques well known to those skilled in the art, and then ligated to purified BamH I-cut arms of coliphage Charon 30. Rimm, D. L, et al, Gene 12:301-309 (1980). Ligation procedures were again those well known to those skilled in the art of recombinant DNA. The T. pallidum DNA-coliphage Charon 30 construct was packaged in vitro, Blattner, F. R., et al., Science 202:1279-1284 (1978) and then used to infect E. coli strain K 802. The resulting plaques were screened for T. pallidum antigens by an in situ radioimmunoassay. Screening was done by a modification of the "Western" blotting procedure of Towbin, H., et al., PNAS USA 76:4350-4354 (1979). Nitrocellulose discs were laid over the phage plaques, and the discs allowed to absorb protein for 10-30 minutes. Little protein was absorbed from unlysed E. coli

of the lawn. The nitrocellulose filters were then coated with ovalbumin by soaking for 10 minutes in 5% ovalbumin in 50 mM tris-HCl (pH 7.5), 150 mM NaCl, 0.15% sodium azide (TSA-5%OA). The plaque blots were incubated overnight in
5 either human secondary syphilitic sera or in normal human sera; both sera were diluted 1:300 in TSA-1%OA. Autoradiograms were prepared as described in Towbin, supra, after the blots were exposed to ¹²⁵I-labelled S. aureus protein A.

One plaque, designated Tp3A, which gave a particularly strong reaction with a secondary syphilitic serum,
10 was used for additional transformations. Phage from plaque Tp3A were diluted and replated on E. coli CSH 18. When rescreened with three different secondary syphilitic sera, all Tp3A plaques produced autoradiograms showing positive
15 radioactive reactions. Autoradiograms from control plaques of cloning vector Charon 30 exhibited little or no radioactivity. This demonstrated that gene products from the Tp3A transformed hosts were antigenic for antibodies in sera of syphilitic individuals.

20 To further study the gene products from the Tp3A transformed hosts, a total protein lysate from the transformed hosts were submitted to SDS polyacrylamide gel electrophoresis as described in Example III. The differentiated polypeptides were then electrophoretically transferred to
25 nitrocellulose strips as described in Example I. The strips were coated with ovalbumin and incubated with syphilitic sera. Again, autoradiograms were prepared as described in Example I after the blots were exposed to ¹²⁵I-labelled S. aureus protein A.

30 FIGURE 19 is a photograph of an autoradiogram of a diagnostic strip of cloned treponemal antigenic peptides from Tp3A transformed hosts. Strip 1 is the differentiated peptide patterns from the transformed hosts following exposure to syphilitic sera. Strip 2 is a Charon 30 control.
35 Strip 3 shows the total T. pallidum protein profile after

exposure to syphilitic sera. Strip 4 is standard system of molecular weight markers.

5 A comparison of strip 1 with strip 4 reveals that Tp3A genes code for at least five peptides of 41,000, 38,000, 23,000, 19,700, and 17,600 molecular weight which react specifically with syphilitic sera. The molecular weights of these cloned antigenic proteins correspond to the molecular weights of antigenic proteins of T. pallidum illustrated: in strip 3. Control strip 2 shows that lysate proteins
10 obtained from Charon 30 transformed hosts do not react with syphilitic sera. This demonstrates that the treponemal antigenic proteins are coded for by the cloned T. pallidum DNA.

15 It may be seen, therefore, that the invention provides in one aspect a method of diagnosing for the presence of a specific disease, a specific disease stage, or allergy in a patient by which a much higher accuracy may be obtained in a very short time. The invention opens the possibility of providing physicians, in their offices or in small laboratories,
20 with the ability to provide quick diagnosis on the basis of a sample of a patient's serum. Long waits and possible inaccuracies which are typical of many widely used clinical diagnosis techniques are eliminated. The technique of the invention is applicable to a wide variety of diseases or
25 allergies, merely requiring an initial series of comparison tests to ascertain and develop the empirical information necessary to select the optimum group of antigenic components and the optimum differentiation process.

30 Various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are intended to fall within the scope of the appended claims. In addition, the invention is not, of course, necessarily limited as an analysis tool to diagnosis.

CLAIMS:

1. A method for producing an analytical antibody probe for detecting the presence of antibodies specific for particular protein and/or polysaccharide material, which
5 method comprises disposing upon a solid-state substrate in a pre-determined spatial relationship a set of differentiated antigenic components which have been derived from said protein or polysaccharide material, said set of differentiated antigenic components being selected such that at least a
10 subset thereof is known to be reactive with said antibodies.

2. A method as claimed in claim 1, wherein the antigenic components are those of a pathogen, an allergen, tumour tissue, or of a virus.

15

3. A method as claimed in claim 1, wherein the antigenic components are proteins, said proteins having been produced as the products of genes derived from an antigenic organism, which genes have been cloned separately into
20 suitable genetically engineered host micro-organisms.

4. A method as claimed in claim 3, wherein the antigenic organism is Treponema Pallidum.

25 5. A method as claimed in any one of claims 1 to 4, wherein the antigenic components are proteins which have been differentiated by means of gel electrophoresis.

30 6. A method as claimed in any one of claims 1 to 4, wherein the antigenic components are proteins which have been differentiated by means of a non-denaturing gel system.

35 7. A method as claimed in claim 6, wherein the non-denaturing gel system is an isoelectricfusing gel system or a native gel system.

8. A method as claimed in any one of claims 5 to 7, wherein after differentiation the antigenic components thus differentiated are transferred from the gel to the solid-state substrate by means of filter affinity transfer.

5

9. An analytical antibody probe for detecting the presence of antibodies specific for particular protein and/or polysaccharide material, which probe comprises a solid-state substrate upon which is disposed in a predetermined spatial relationship a set of differentiated antigenic components derived from said protein or polysaccharide material, said set of differentiated antigenic components being selected such that at least a subset thereof is known to be reactive with said antibodies.

10

10. A probe as claimed in claim 9 further defined by the feature or features of any one or more of claims 2 to 8.

15

11. A method for analysing a sample to determine the presence and distribution of antibodies specific for particular protein and/or polysaccharide material, which method comprises providing a solid-state substrate upon which is disposed in a predetermined spatial relationship a set of differentiated antigenic components derived from said protein or polysaccharide material, said set of differentiated antigenic components being selected such that at least a subset thereof is known to be reactive with said antibodies, contacting said substrate with said sample under conditions which permit reaction of any of said antibodies in the sample with the antigenic components on the substrate, and detecting the existence and pattern of antigen-antibody reactions on the substrate.

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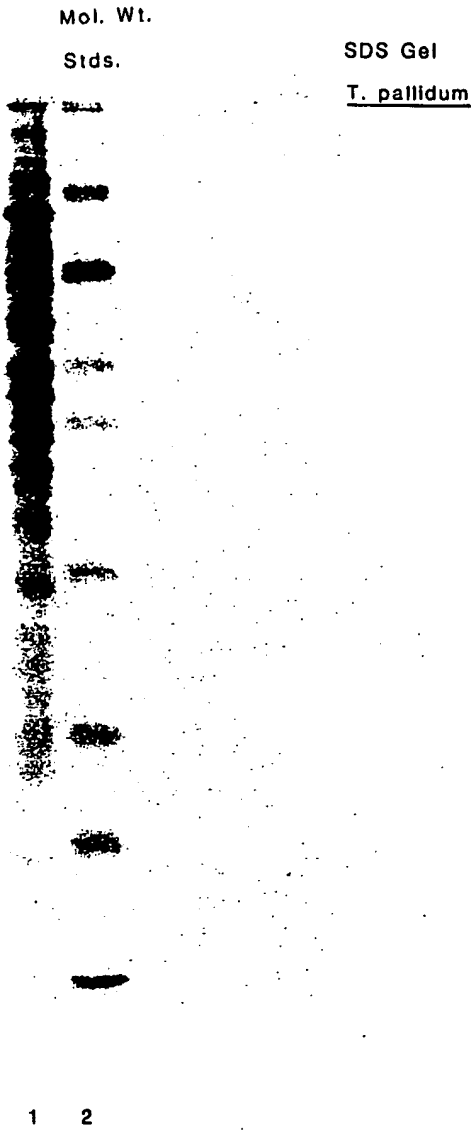


FIG. 1

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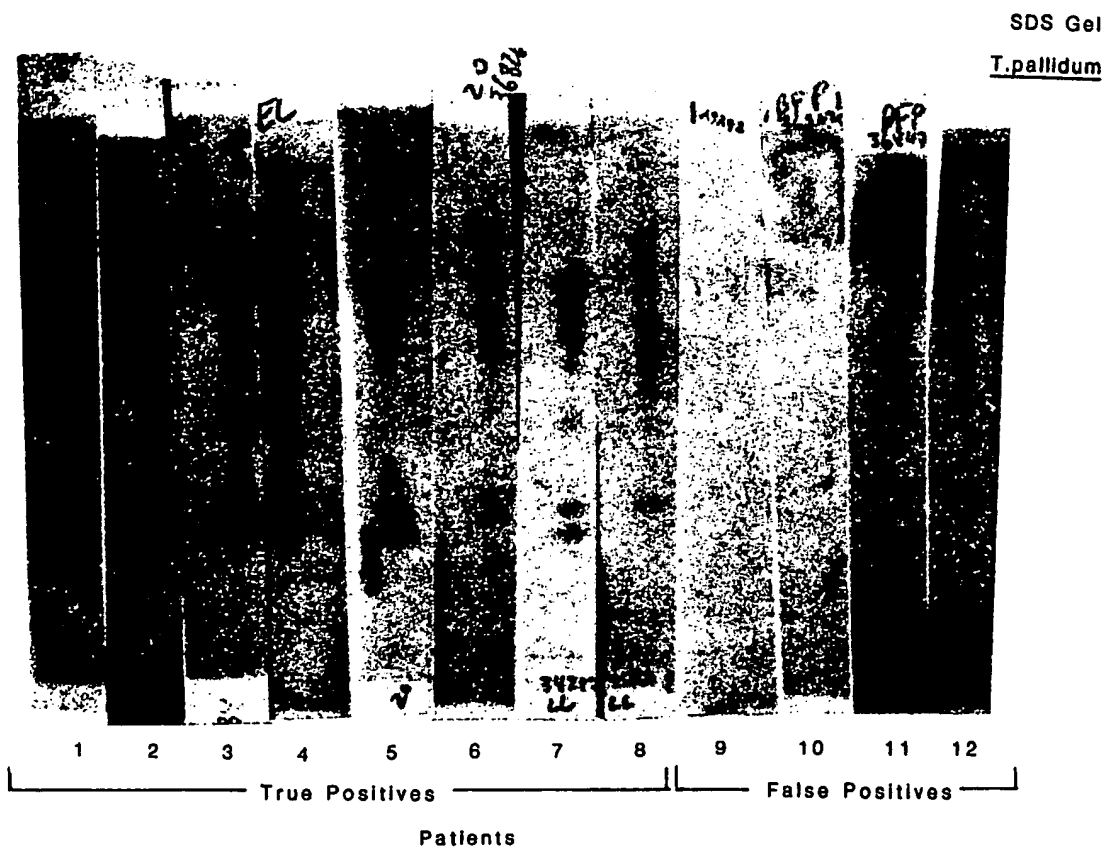


FIG. 2

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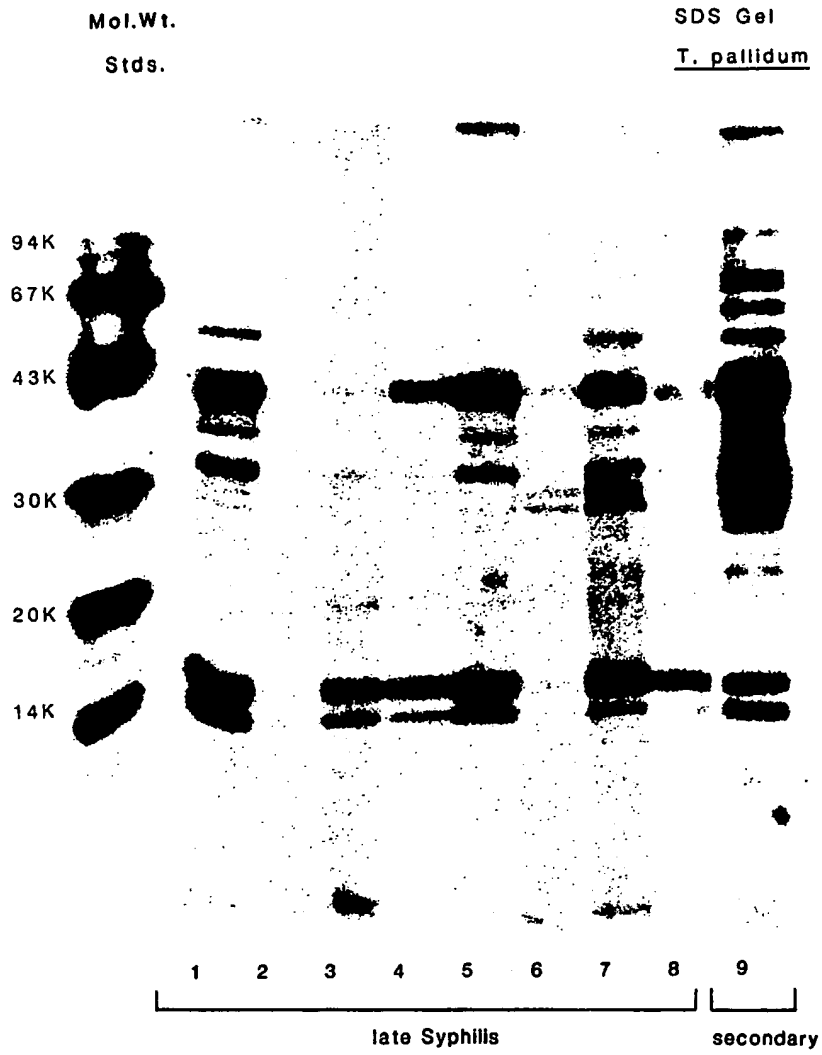


FIG. 3

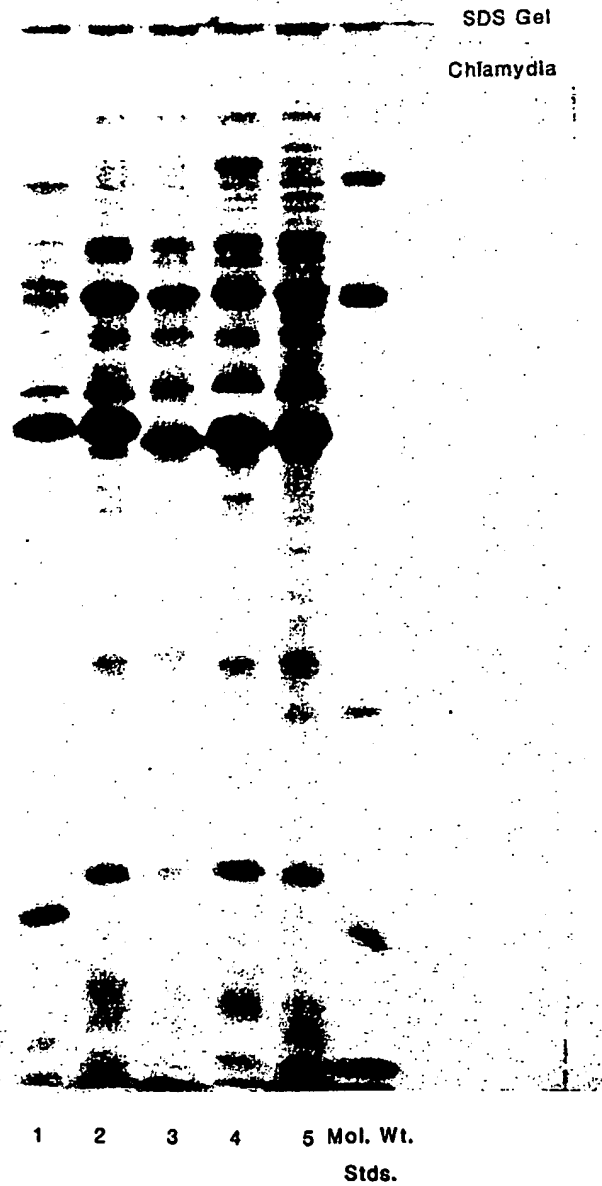
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FIG. 4

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FIG. 5



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FIG. 6

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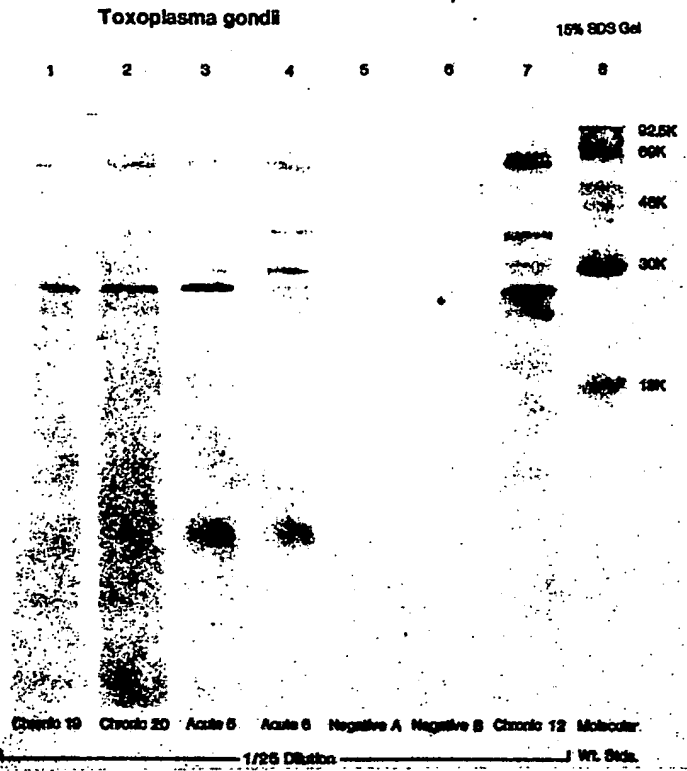


FIG. 7

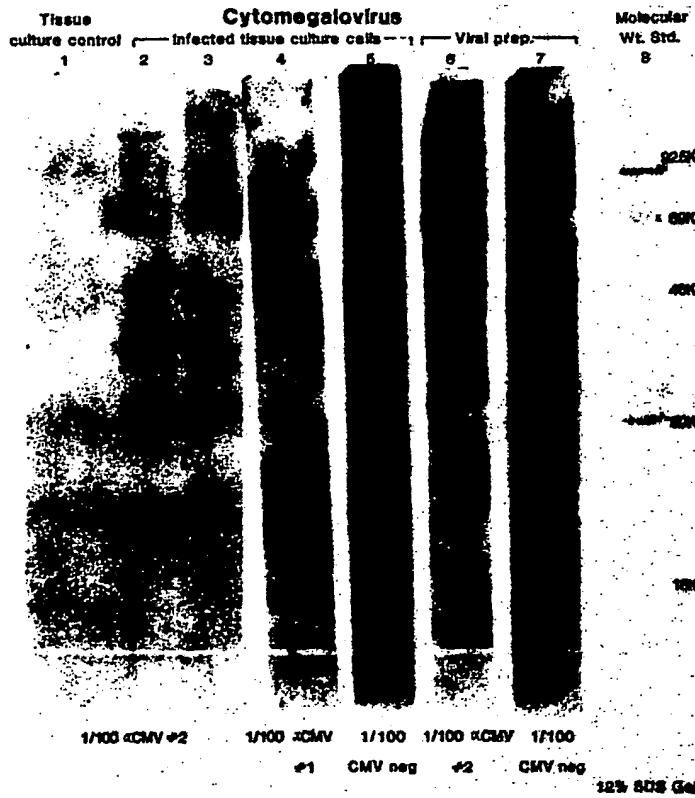


FIG. 8

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FIG. 9

Streptococcus pneumoniae

Type 4

Type 8

Normal

Immune

FIG. 10

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L₂ C. trachomatis
type D infection

SDS Gel
bee venom

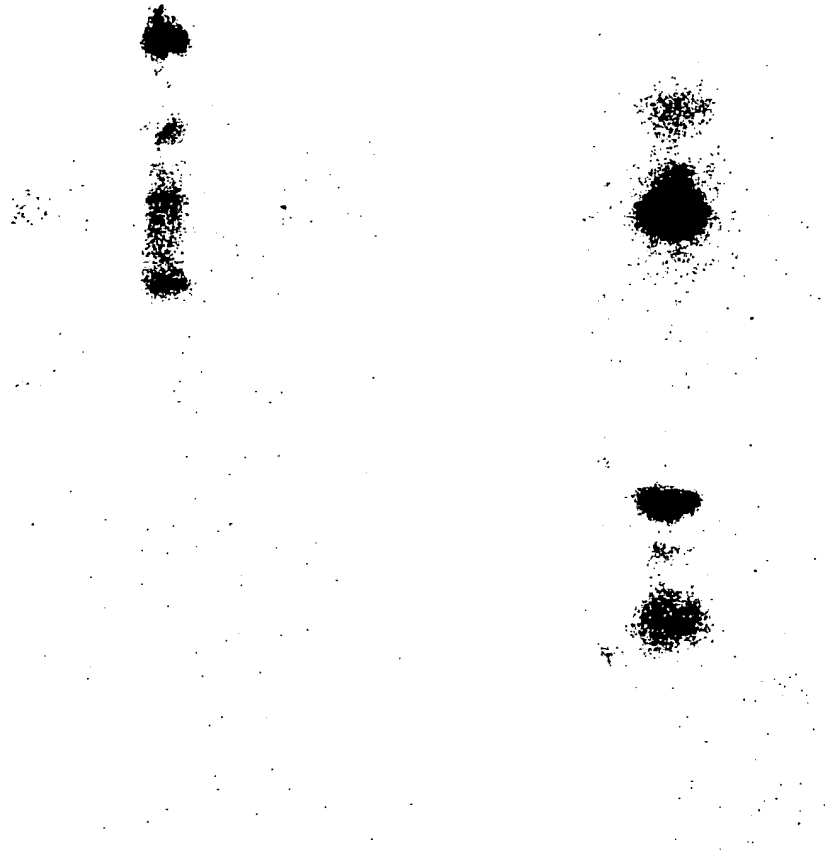


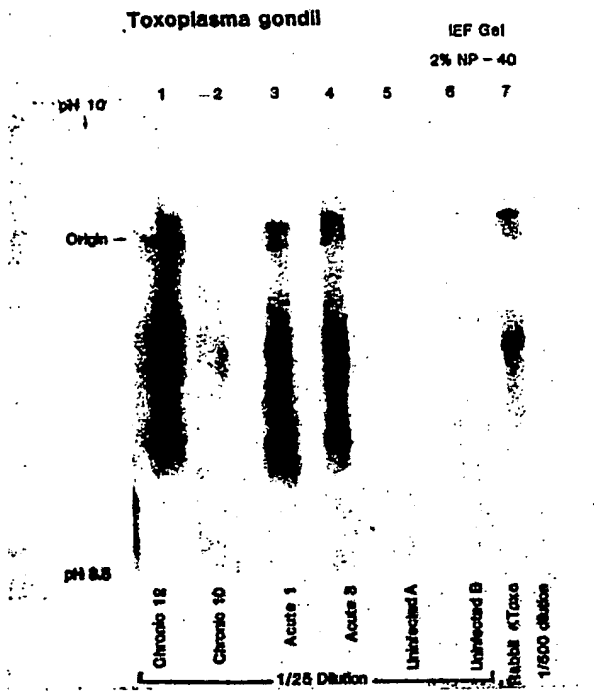
FIG. 13

FIG. 11

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FIG. 17



L₁ C. trachomatis
type F infection

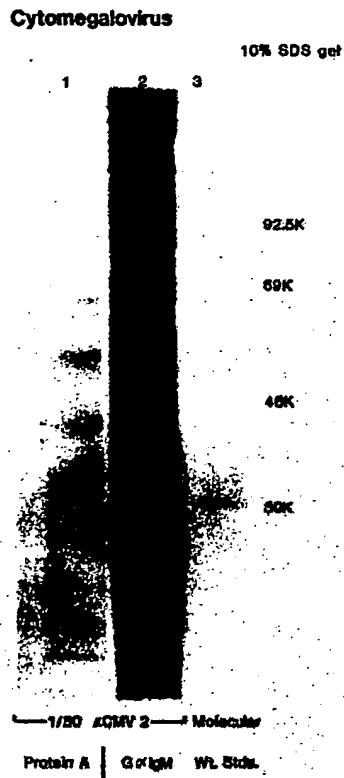


FIG. 12

FIG. 16

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FIG. 18



Cloned *T. pallidum*

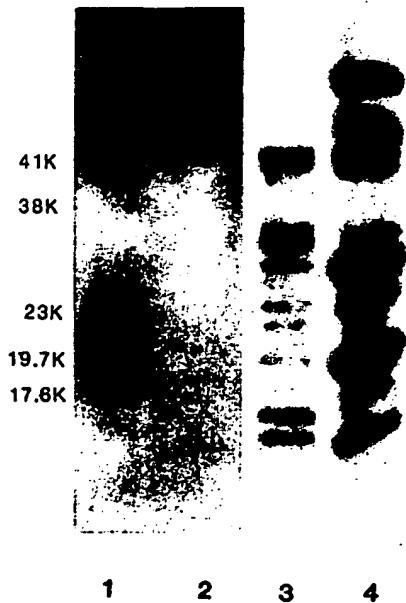


FIG. 19



DOCUMENTS CONSIDERED TO BE RELEVANT		CLASSIFICATION OF THE APPLICATION (Int. Cl. 3)	
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	
D	<p>PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCE, vol. 76, no. 9, September 1979 USA H. TOWBIN et al.: "Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: Procedure and some applications" pages 4350-4354 * The whole article *</p> <p>---</p> <p>GB - A - 2 008 747 (INTERNATIONAL DIAGNOSTIC TECHNOLOGY) * Page 2, line 1 - page 3, line 12; claims 1 & 5-10 *</p> <p>---</p> <p>E EP - A - 0 017 460 (RESEARCH CORP.) * Page 11, line 14 - page 20; claims 1 and 2 *</p> <p>---</p> <p>D CHEMICAL ABSTRACTS, vol. 92, no. 7, 18 February 1980, page 320, column 1, abstract no. 54615x COLUMBUS, OHIO (US) & J. BIOL. CHEM., vol. 254, no. 23, 1979, pages 12240-12247 H.A. ERLICH et al.: "Filter affinity transfer: a new technique for the in situ identification of proteins in gels" * The whole abstract *</p> <p>---</p> <p>./..</p>	<p>1,2,5- 7,9- 11</p> <p>1,2,4 9-11</p> <p>1,2,9- 11</p> <p>1,8</p>	<p>G 01 N 33/54 27/26</p> <p>TECHNICAL FIELDS SEARCHED (Int.Cl. 3)</p> <p>G 01 N 33/54 33/52 27/26</p> <p>CATEGORY OF CITED DOCUMENTS</p> <p>X: particularly relevant if taken alone Y: particularly relevant if combined with another document of the same category A: technological background O: non-written disclosure P: intermediate document T: theory or principle underlying the invention E: earlier patent document, but published on, or after the filing date D: document cited in the application L: document cited for other reasons</p> <p>&: member of the same patent family, corresponding document</p>
<p>A The present search report has been drawn up for all claims</p>			
Place of search	Date of completion of the search	Examiner	
The Hague	25-01-1982	GRIFFITH	



DOCUMENTS CONSIDERED TO BE RELEVANT			CLASSIFICATION OF THE APPLICATION (Int. Cl.)
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	
	<p>CHEMICAL ABSTRACTS, vol. 92, no. 1, 7 January 1980, page 276, column 1, abstract no. 2752v COLUMBUS, OHIO (US) & SEIBUTSU BUTSURI KAGAKU, vol. 22, no. 4, 1979, pages 279-284 N. TAKAHASHI et al.: "Immunochemical detection of plasma proteins after two-dimensional electrophoresis"</p> <p>* The whole abstract *</p> <p>---</p>	1,5-7, 9-11	
	<p>CHEMICAL ABSTRACTS, vol. 73, no. 11 September 14, 1970, page 66, column 2, abstract no. 52950m COLUMBUS, OHIO (US) & CLIN. CHIM. ACTA, vol. 28, no. 1 1970, pages 149-152 J.H. DEWAR et al.: "Immunological technique for identifying protein areas in gel slabs"</p> <p>* The whole abstract *</p> <p>---</p>	1,5-7 9,10	TECHNICAL FIELDS SEARCHED (Int. Cl.)
	<p><u>US - A - 3 941 876</u> (V.A. MARINKOVICH)</p> <p>* Column 1, lines 45-62; column 5, lines 39-46; claims 1-3 *</p> <p>---</p>	1,2,9- 11	
A	<p><u>US - A - 4 094 759</u> (G. RUHENSTROTH-BAUER et al.)</p>		
A	<p><u>US - A - 4 097 149</u> (F.J. ALADJEM et al.)</p>		
A	<p><u>FR - A - 2 233 024</u> (GREYWARD LABORATORIES. INTERCONTINENTAL LTD.)</p>		
A	<p><u>US - A - 3 979 509</u> (I. GIAEVER)</p> <p>-----</p>		